

Human Induced Pluripotent Stem Cells (from Normal, Female, Peripheral Blood Mononuclear Cells)

Control Human Induced Pluripotent Stem cells (iPSCs) at low passage (p8) provided by Applied StemCell Inc. This iPSC line was generated from normal Peripheral Blood Mononuclear Cells (PBMCs) using a Sendai virus method. This method allows the expression of human transcription factors (Oct-4, Sox-2, KLF4, and c-MYC) that initiate the reprogramming process. The resulting induced pluripotent cells were selected using morphological criteria without the use of either fluorescent markers or drug selection.

Catalog #	Product	Size
ASE-9217	Human iPSCs (from Normal, Female, PBMCs)	1.0 × 10 ⁶ cells/vial

Parental Tissue Information	
Reprogramming Method	Sendai Virus
Passage Number	p8
Tissue	Peripheral Blood Mononuclear Cells (PBMCs)
Sex	Female
Age	31
Race	Caucasian
Clinical Information	Normal

Intended Use

The ASE-9217 control human induced pluripotent cell line can be used for CRISPR-CAS9 genome editing and differentiation into somatic lineages *in vitro*.

This product is for research use only and not intended for human or animal diagnostic or therapeutic uses.

Quality Assurance

Each lot of ASE-9217 human iPSCs has been tested for viability following recovery from cryopreservation, morphology, expression of pluripotency markers (Oct-4, Sox-2, and SSEA-4) and for normal female karyotype. Additionally, these iPSCs have been tested to confirm the absence of bacterial, fungal, and mycoplasma contamination.

All quality control test results are reported on a lot-specific Certificate of Analysis, available at appliedstemcell.com or upon request.

Storage Instructions and Stability

Store in liquid nitrogen freezer immediately upon receipt.

For shelf-life data check lot specific Certificate of Analysis.

Precautions

The safety and efficacy of this product in diagnostic or other clinical uses have not been established. This product is for research use only and not intended for human or animal diagnostic or therapeutic uses.

Please be aware that the following scenario can occur: Liquid nitrogen can leak into the vials while they are submerged in a liquid nitrogen freezer. During rapid thawing, the liquid nitrogen returns to the gas phase, resulting in a dangerous build-up of pressure within the vial. This can result in the vial exploding and expelling not only the vial contents but also the vial cap and plastic fragments of the vial. When retrieving the cell vial from a liquid nitrogen freezer, carefully twist open the cap to release any built-up nitrogen gas prior to thawing the cells.

Refer to the Safety Data Sheet (SDS) for information regarding hazards and safe handling practices.

Directions for Use

Required Materials (Not Provided)

Product	Vendor	Catalog#
mTeSR-Plus Medium	StemCell Technologies	05825
(Optional)100 µg/mL Primocin™	Invivogen	ant-pm-2
Matrigel®	Corning	354277
Geltrax	ThermoFisher Scientific	A1413202
ROCK Inhibitor (Y-27632)	Sigma-Aldrich	SCM075
CloneR	StemCell Technologies	05889
CryoStor® CS10	StemCell Technologies	7930
0.5 M EDTA in PBS	ThermoFisher Scientific	15575-020
PBS	Life Technologies	14190136
Cell Scraper	VWR International	75799-938
Corning® CoolCell® FTS30	Corning®	432006

Protocol for Thawing human iPSCs

1. Prepare Matrigel® or Geltrex coated 6-well plates in advance, following vendor's instructions.
Note: Prepare Matrigel or Geltrex plates the day before and not more than 3 days prior to thawing. Before transferring the cells, the Matrigel must be aspirated.
2. Prepare 6 mL of mTeSR-plus medium (mix of 4.8 mL basal medium and 1.2 mL 5x supplement) + Primocin (1:500 dilution) + 10 µM Rock Inhibitor.
Note: Besides Rock inhibitor, the clone R can be used.
3. Add 1 mL of mTeSR-plus medium + Rock Inhibitor + Primocin in one well of a Matrigel®-coated plate. Prepare 1 well for each vial of frozen cells.
4. Bring the cryovial on dry ice to the tissue culture room.
5. Quickly thaw the iPSCs in a 37°C water bath, by gently shaking the cryovial continuously until only a small piece of ice remains.
6. Wipe the cryovial with a paper towel sprayed with 70% ethanol and place it into a biosafety cabinet.
7. Add 9 mL of mTeSR-plus medium + Primocin to a 15 mL conical tube.
8. Using a 1 mL pipette transfer the cells to the 15 mL conical tube dropwise while swirling the conical tube.
9. Centrifuge the cells for 3 minutes at 200 RCF and at room temperature.
10. Aspirate off the medium and add 3 mL mTeSR-plus medium + Rock Inhibitor + Primocin.
11. Gently flick the conical tube to resuspend the cells and transfer them to the 1 well of the Matrigel® or Geltrex plate using a 5 mL serological pipette.
12. Place the plate in the incubator and move the plate back-and-forth and side-to-side, twice to spread the clumps evenly in the wells.
13. Medium can be changed 2 days after thawing. After the 2 days, if the number of attached colonies is still low (less than 5% confluency), change half of the medium (aspirate 1 mL and add 1 mL of mTeSR-plus medium + Primocin).
14. Once colonies are stabilized, change the medium daily. Usually, within 1 week the cells are ready to be split.

15. When the ASE-9217 hiPSC colonies are big or close enough to merge, the cells need splitting/passaging.

Protocol for Passaging and Seeding human iPSCs

1. Aspirate the medium from the hiPSC culture.
2. Wash once with 1 mL of 0.5 mM EDTA (in PBS).
3. Aspirate the EDTA and add 1 mL of 0.5 mM EDTA (in PBS) per well of a 6-well plate and incubate the cells for 3 min in a 37°C incubator.
4. Observe the cells under a microscope until the cells at the edge of the colonies start to separate and round up.
5. With the cells still attached, aspirate the EDTA and add 1 mL of mTeSR-plus medium.
6. Scrape the cells from the bottom of the well until the colonies are floating; pipette up and down 2-3 times (with the 1000 µL pipette set to 800 µL) to break the colonies in small clumps.

Note: Pipetting up and down three times is enough to break the colonies into clumps of optimal size.

7. Transfer the desired dilution to the wells of the new Matrigel®-coated plate (usually around a 1:3 splitting ratio).

Note: Prepare Matrigel plates (or any other basement matrix) not more than a week prior to passaging cells.

8. Place the plate in the incubator and move the plate back and forth and side to side, twice to spread the clumps evenly in all the wells.

Note: Human iPSCs are passaged as clumps of 50-200 cells, rather than single cells. Cells need to be passaged before the colonies are large enough to merge with one another.

Protocol for Freezing human iPSCs

1. Label the cryovial as needed, based on 1 vial per well of a 6-well plate, and pre-chill them in a 4°C freezer.
2. Aspirate the medium from the hiPSC culture.
3. Wash once with 1 mL of PBS.
4. Aspirate the PBS and add 1 mL of 0.5 mM EDTA (in PBS) per well of a 6-well plate and incubate the cells for 3-5 minutes in a 37°C incubator.
5. Observe the cells under a microscope. After 1-2 minutes the cells at the edge of the colonies will start to separate and round up.
6. Once round colonies are observed, aspirate the EDTA (even if it hasn't been 5 minutes of incubation).
7. Add 1 mL of cold freezing medium, CS10 and scrape the cells from the bottom of the well until the colonies are floating.

Note: The freezing medium must remain at 4°C until usage.

8. Pipette the cells once or twice to break up any big clumps before transferring the suspension to a cryovial.
9. Transfer the cell suspension into the pre-chilled and labeled cryovial.
10. Place the cryovial in a CoolCell® Freezing Container or in a Styrofoam rack at -80°C overnight, and transfer to liquid nitrogen the next day.

Data

Morphology Analysis

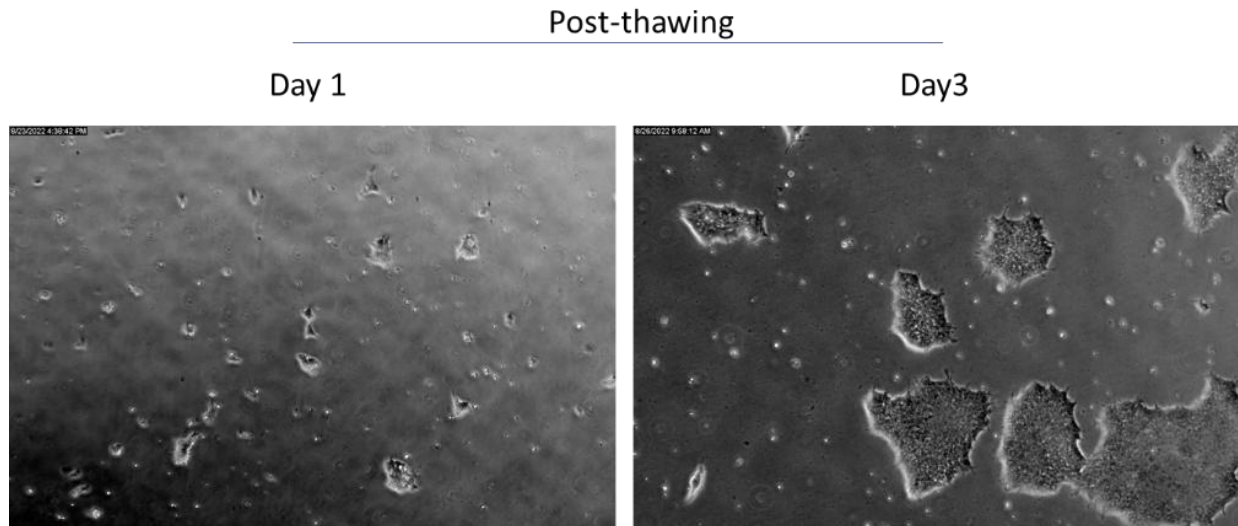


Figure 1. Morphology images of the ASE-9217 iPSC line captured post thaw (Day 1 and 3). The control human iPSCs were generated from normal PBMCs using a Sendai virus method. The image on the left was taken on day 1 and the image on the right was captured day 3.

Pluripotency Marker Analysis

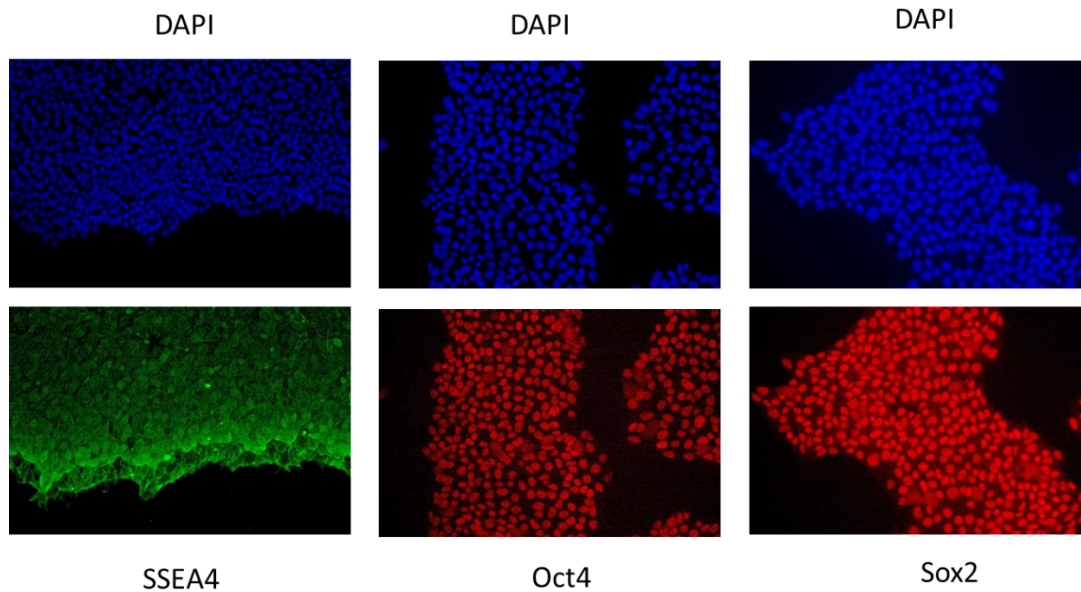


Figure 2. Expression of pluripotency markers. ASE-9217 iPS cell line expresses common iPSC biomarkers (Bottom row: *SSEA4*, *Oct4*, and *Sox2*). Top row: Corresponding DAPI nuclear staining.

Karyotype Analysis

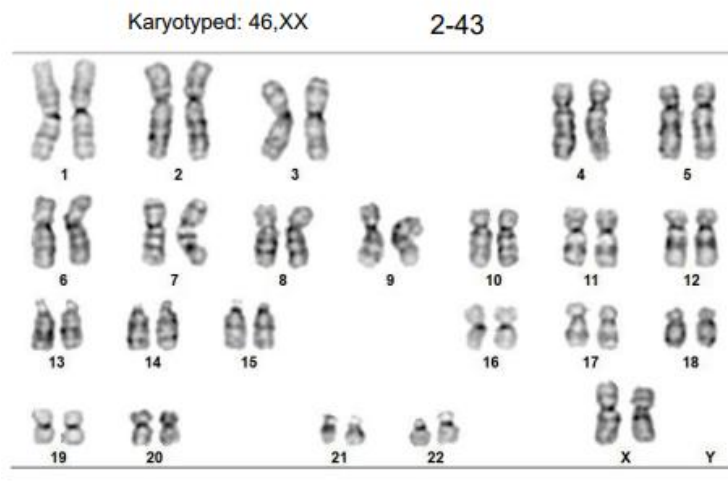


Figure 3. Karyotype analysis to rule out genetic aberrations. The G-banded chromosome analysis of metaphase ASE-9217 cells demonstrate a normal female karyotype.

Directed Differentiation into Germ Layers

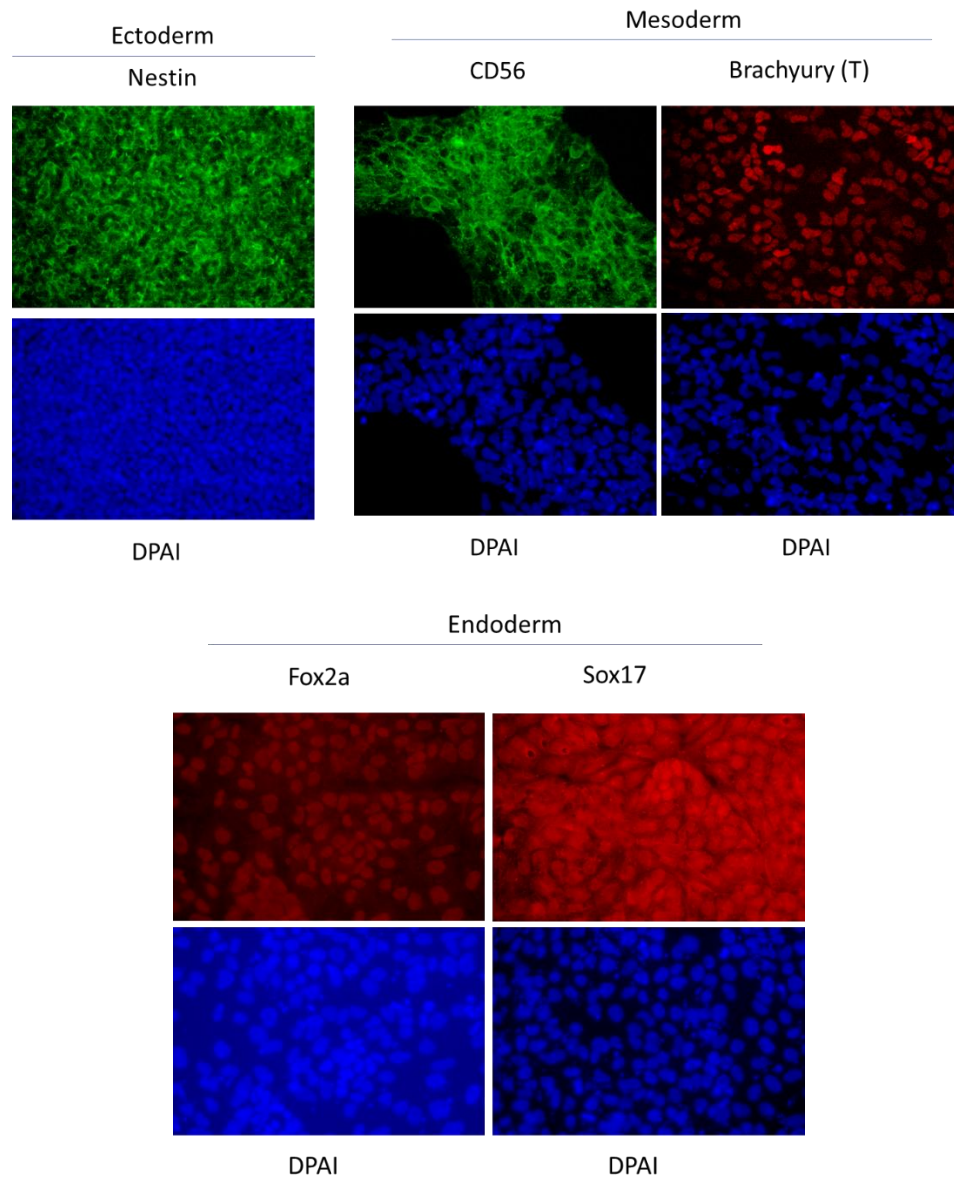


Figure 4. Direct differentiation of ASE-9217 to three germ layers. The ASE-9217 hiPSC line was differentiated to specific lineages of the germ layers using well-established and optimized protocols. Immunostaining for biomarkers of each lineage was performed to confirm lineage commitment. Cells were also co-stained with nuclear marker, DAPI (blue). Ectoderm (EC) marker: Nestin (green); Mesoderm (ME) markers: CD56 (green) and Brachyury (T) (red); Endoderm (EN) markers: Fox2a (red) and Sox17 (red).

Related Products

Catalog #	Product	Size
ASE-9215	Human iPSCs from PBMCs (Normal, Male, Caucasian, Adult)	1 X 10 ⁶ cells/vial
ASE-9209	Human iPSCs from Fibroblasts (Normal, Female, Caucasian, Adult)	1 X 10 ⁶ cells/vial
ASE-9216	Human iPSCs from Dermal Fibroblasts (Normal, Male, Caucasian, Neonate)	1 X 10 ⁶ cells/vial
ASE-9211	Human iPSCs from Fibroblasts (Normal, Male, African American, Neonate, NIST line)	1 X 10 ⁶ cells/vial

Technical Support

CONTACT US

For more information or assistance, contact Customer Service at:

- Email: info@appliedstemcell.com
- Direct line: [\(408\) 773-8007](tel:4087738007)

WEBSITE RESOURCES

Visit appliedstemcell.com for technical resources and information including:

- Safety Data Sheets (SDS)
- Certificates of Analysis (when available)
- FAQs
- Product literature
- Complete contact information

Applied StemCell, Inc.

521 Cottonwood Dr, Suite # 111, Milpitas, California, 95035, USA • Telephone: +1 866.497.4180 • appliedstemcell.com

© 2026 Applied StemCell, Inc. All rights reserved. TARGATT™ and ActiCells™ are trademarks of Applied StemCell, Inc. All other trademarks are the property of their respective owners. PN: IFU-ASE-9217 Rev. 1

Effective Date: 15 September 2026