

Datasheet

SNL 76/7 Feeder Cells, P12, Untreated

Product Information

Specifications

| Catalog Number | Cells per Vial | Treatment | Number of Vials |
|----------------|---------------------|-----------|-----------------|
| ASF-1305 | 2 x 10 ⁶ | Untreated | 1 |

Description

Mouse embryonic stem cells (ESCs) and induce pluripotent stem cells (iPSCs) generally require culture on feeder cells as well as medium containing Leukemia Inhibitory Factor (LIF) to maintain pluripotency and the capacity to self-renew. The SNL 76/7 cell line, derived from a mouse fibroblast STO cell line transformed with neomycin resistance and murine LIF genes, can be used as feeder cells for mouse and human ESC and iPSC culture. The SNL cells are sensitive to HAT selection (hypoxanthine, aminoprotein and thymidine) and negative for HPRT (hypoxanthine quanine phosphoribosyl transferase). Before use, SNL 76/7 cells must be mitotically

inactivated by y-irradiation or mitomycin-C treatment.

P12 **Passage**

Untreated Treatment

Dry ice Shipping

Storage and Stability Store in liquid nitrogen freezer immediately upon receipt. This product is stable for at

least 6 months from the date of receiving when stored as directed.

Biosafety Level BSL-1

Safety Precaution PLEASE READ BEFORE HANDLING ANY FROZEN VIALS. Please wear the

appropriate Personal Protection Equipment (lab coat, thermal gloves, safety goggles and a face shield) when handling the cells. Handle the frozen vials with due caution. Please be aware that the following scenario can occur: Liquid nitrogen can leak into the vials when the vials are submerged in liquid nitrogen. Upon thawing, the liquid nitrogen returns to the gas phase, resulting in a dangerous build-up of pressure within the vial. This can result in the vial exploding and expelling not only the vial

contents but also the vial cap and plastic fragments of the vial.

This product is for research use only and not intended for human or animal diagnostic or **Restricted Use**

therapeutic uses.

Media and Material

Medium

| Component | Concentration | Vendor |
|--------------------------|---------------|-------------------------------|
| ESC-Sure™ DMEM | | Applied StemCell, #ASM-5001 |
| ESC-Sure™ FBS | 10% | Applied StemCell, #ASM-5007 |
| Nonessential amino acids | 0.1 mM | Life Technologies, #11140-050 |
| Sodium Pyruvate | 1 mM | Life Technologies, #11360-070 |
| L-Glutamine | 2 mM | Life Technologies, #25030-164 |

Suggested Neomycin (G418) concentration: 200 µg/mL

Protocol

- Remove a vial of frozen cells from liquid nitrogen and place it onto dry ice for 5 minutes before thawing it at 37 °C
 water bath. As soon as the majority of the content of the vial thawed, transfer it to a conical tube containing 10x
 volume of pre-warmed medium.
- 2. Spin at 1000 rpm for 5 minutes at room temperature, discard medium, resuspend the cells in growth medium and plate them at an appropriate density in a gelatin-coated tissue-culture dish or flask (25,000-50,000 cells/cm²). Optimal density is to be determined by the user for specific applications.

Tip: make sure coating with 0.1-0.2% Gelatin solution in 37 degree incubator for at least 30 minutes.

- 3. After 2-3 days (cells should have grown to a density of ~150,000 cells/cm²), trypsinize the cells and subculture at 1:3 ratio.
- 4. For use as feeder cells, plate mitotically inactivated cells (see below) at an appropriate density in a gelatin-coated tissue-culture dish or flask (generally 75,000 cells/cm². Optimal density is to be determined by the user for specific applications.
- Mitotic inactivation by y-irradiation

When cells reach confluency, trypsinize the cells, spin down, resuspend cells in chilled growth medium, and γ-irradiate the cell suspension at 3000 rad.

Mitotic inactivation by Mitomycin-C treatment

When cells are confluent, treat the cells with 10 µg/ml mitomycin C for 2 hours, then trypsinize the cells, spin down and resuspend in growth medium and plate for use (cells can also be frozen down in freezing medium).

References

- 1. McMahon, A.P. and Bradley, A. (1990) Cell 62:1073-1085.
- 2. Okita, K; Ichisaka, T; Yamanaka, S. (2007) Nature 448:313–317.
- 3. Takahashi K, Okita K, Nakagawa M, Yamanaka S. (2007) Nat Protoc. 2:3081-9.
- 4. Takahashi K, Narita M, Yokura M, Ichisaka T, Yamanaka S. (2009) PLoS One 4(12):e8067.